

larvi 2013

6th fish & shellfish larviculture symposium





Reproduction of European eel and larval culture State of the art





ghent university, belgium, 2-5 september 2013





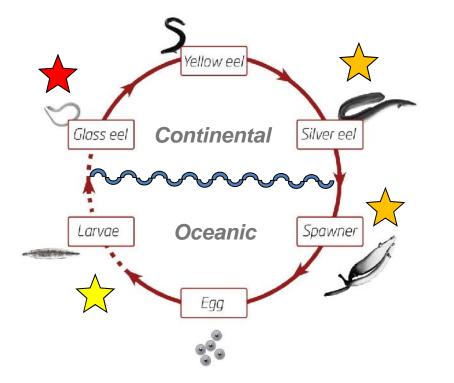
Reproduction of European eel and larval culture - State of the art

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- Eel aquaculture relies on wild caught glass eels
- Severe decline in eel stocks renders present aquaculture production unsustainable
- Captive breeding is needed to sustain eel aquaculture



Challenges for hatchery production of glass eels

Complex life cycle

Hormonal control and inhibition of maturation during silvering

Lack of knowledge about wild eel reproduction and early life stages

PRO-EEL OBJECTIVES



Increase knowledge about eel physiology and hormonal control of reproduction

Improvement of broodstock nutrition and hormonal treatments for induction of maturation

Uncover mechanisms of final oocyte maturation, sperm function and fertilisation capability

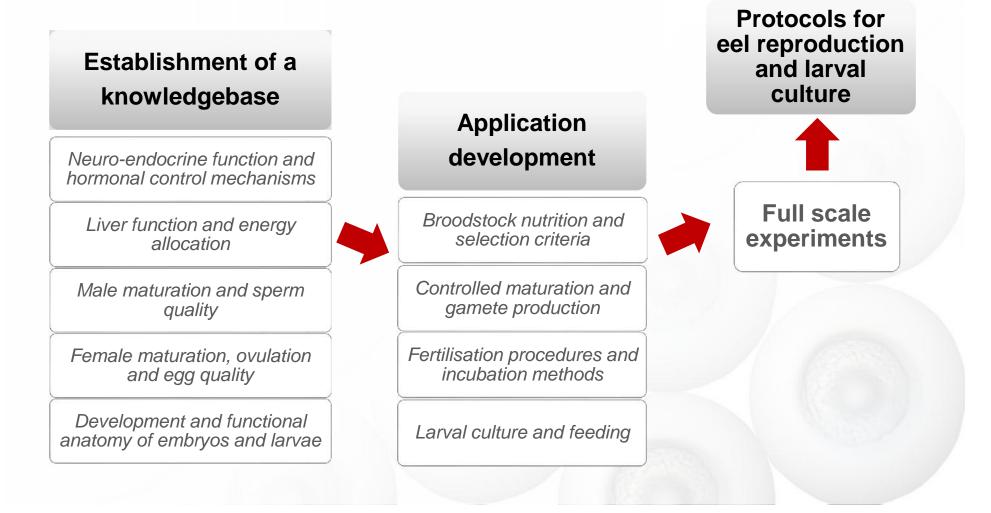
Establishment of a stable production of viable eggs and embryos

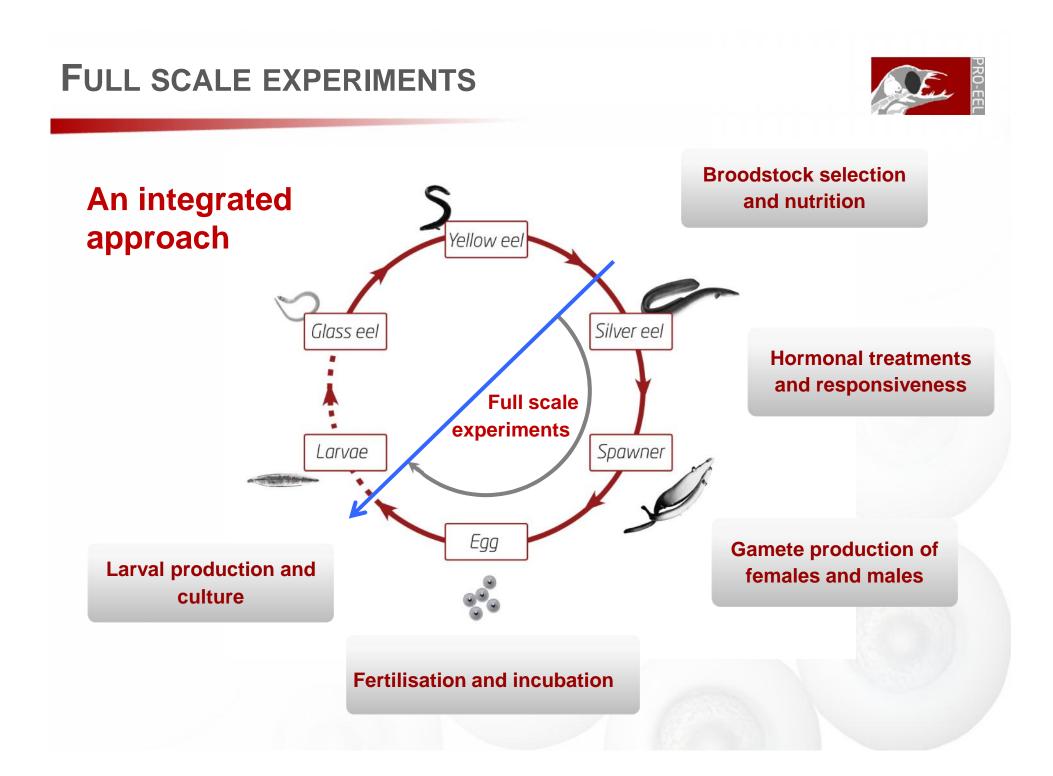
Access requirements for successful egg and larval development and identify suitable larval feeds

Establishment of rearing technology and first feeding in culture of European eel larvae



Link basic research to application and technology development to fill gaps in knowledge and methodology





FULL SCALE EXPERIMENTS



Comparison of reproductive success of farmed and wild broodstocks

Protocols

Broodstock maturation – Wild and farmed: different diets and hormonal treatment

Fertilisation (4-5 males per female) and incubation

Hatching and larval rearing improving gradually technology

Results

Broodstock response Stripping success

Fertilisation success Embryonic developmental success

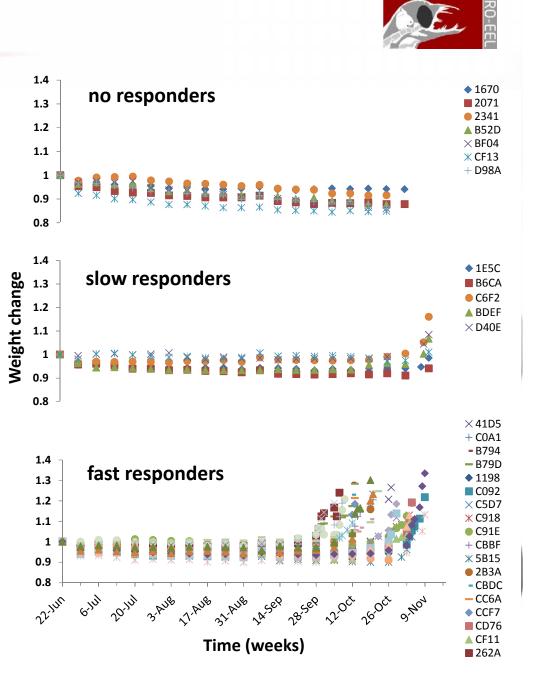
Larval production success Larval quality and longevity

BROODSTOCK RESPONSE

Female response to hormonal treatment varies

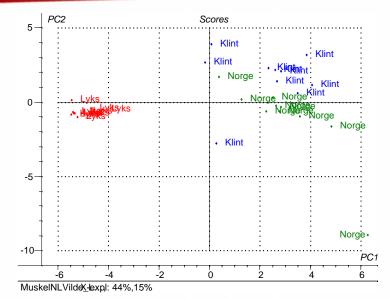
Response of females as weight change over time classified by ovarian histology at sacrifice





FEMALE BROODSTOCK NUTRITION



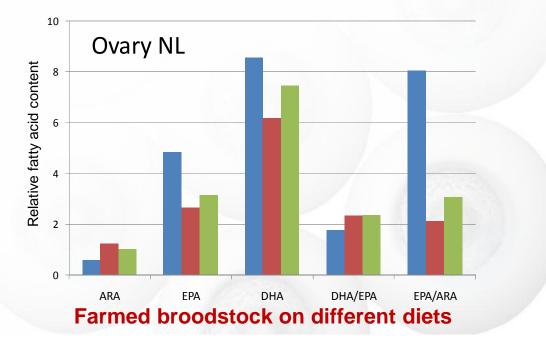


	DAN-EX	JD1	JD2
Protein %	48.1	47.3	47.4
Fat %	25.7	22.9	24.6
EPA g/kg	19.4	8.1	11.9
DHA g/kg	24.4	17.2	20.5
ARA g/kg	1.08	4.51	3.12
Vit. C ppm	257	257	400
Vit. E ppm	163	163	400

Essential FA in different diets

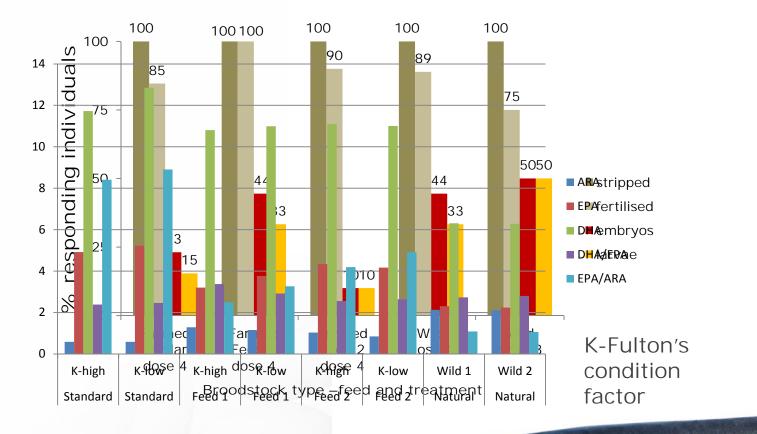
Enhanced farmed broodstock nutrition optimising egg & larvae quality

- Comparing farmed and wild eels
- Formulated diets





Comparing performance of farmed and wild broodstocks

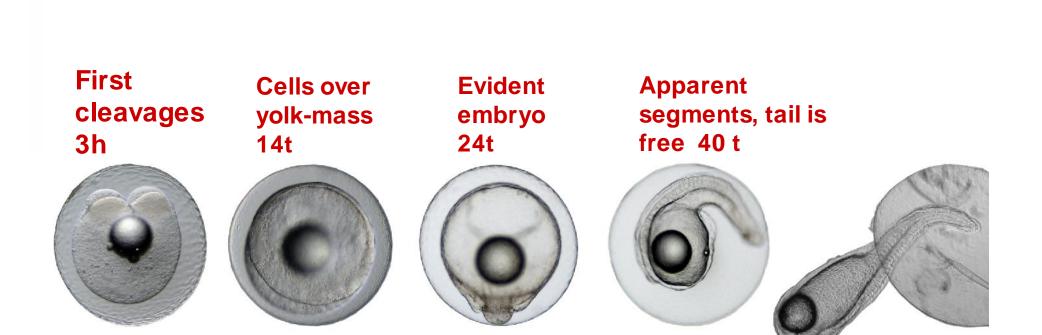




Does feed matter – Yes!

FWRTILISATION AND EMBRYONIC DEVELOPMENT

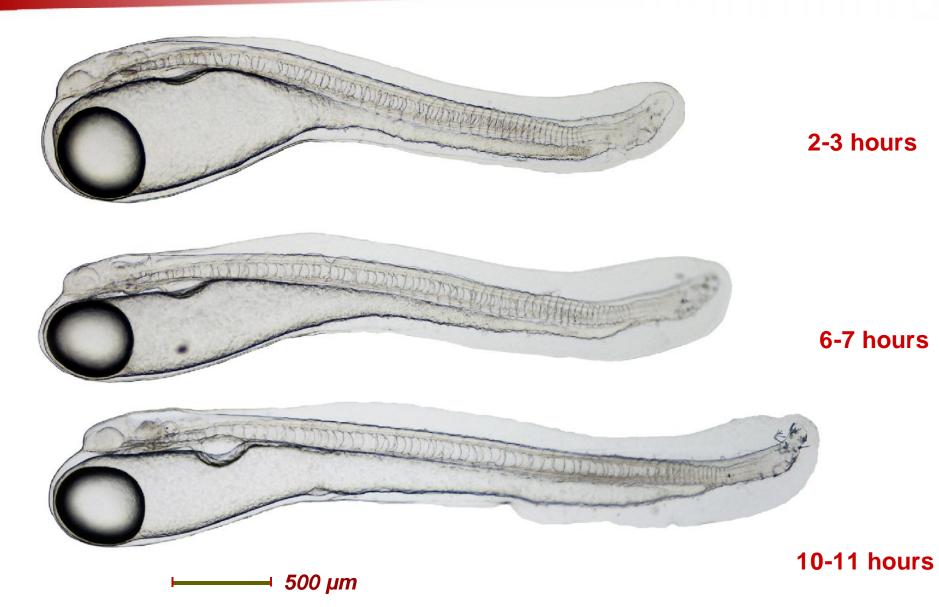


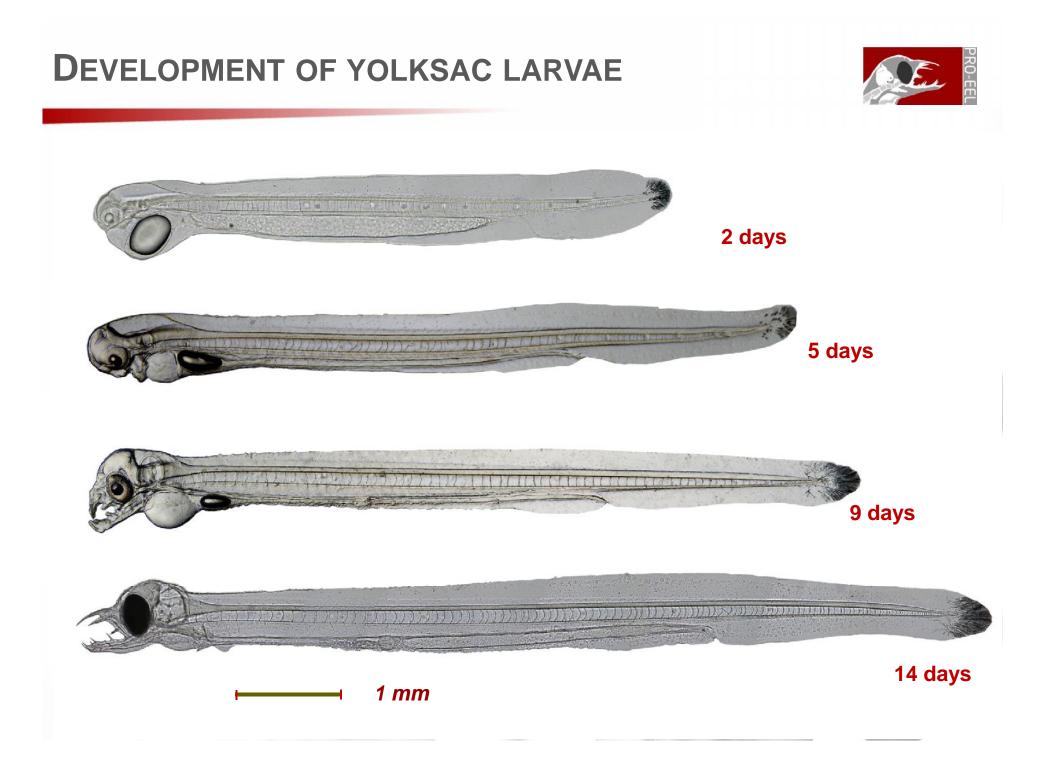


Embryonic stage duration app. 48 h until hatch

DEVELOPMENT OF YOLKSAC LARVAE



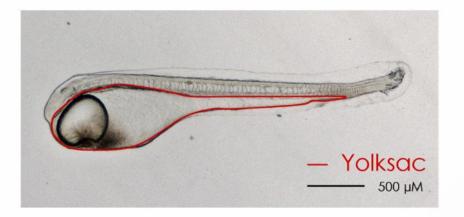




QUALITY OF YOLKSAC LARVAE

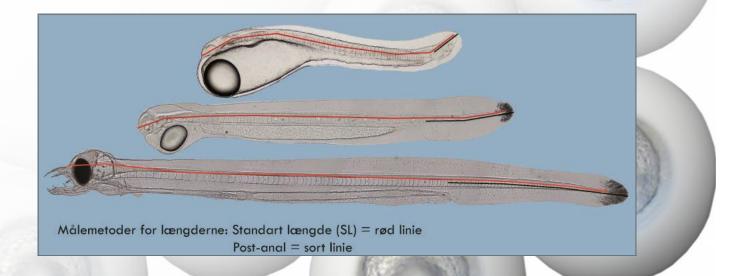


Measurments of morphological characteristics and general developmental success



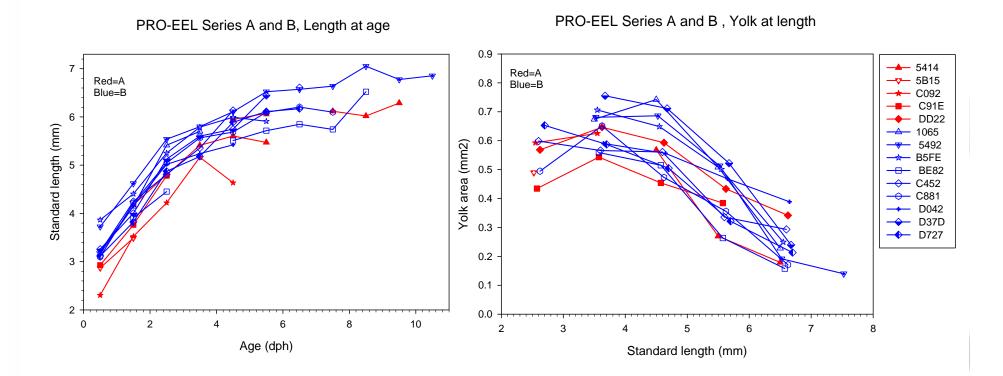
Amount and use of yolk

Length increase



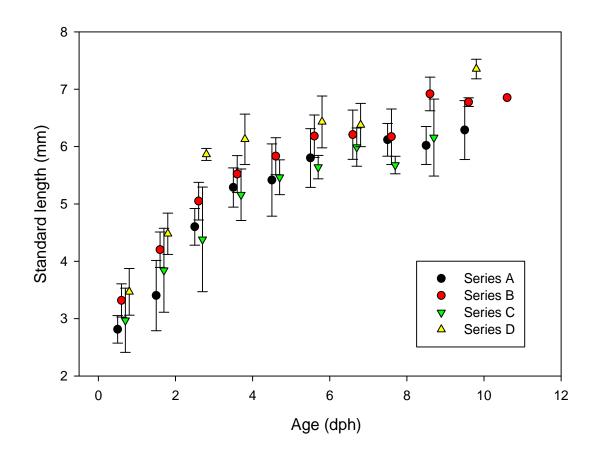


Measuring development in general morphology, amount and use of yolk and length increase





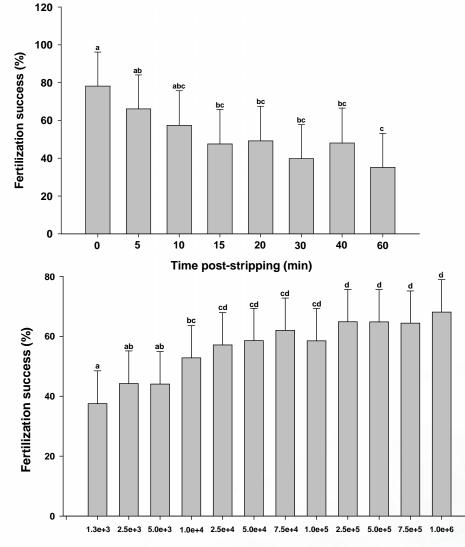
Differences in larval quality between farmed and wild broodstock



Differences in larval quality Farmed: Series: A and C Wild: Series B and D

STANDARDISATION OF FERTILISATION PROCEDURES





Sperm to egg ratio

Standardisation of fertilisation procedures

Importance of sperm to egg ratio and time post-stripping

- Egg fertilisation capacity in relation to time poststripping
- Optiomal sperm # and time post-stripping
 - –25,000 sperm per egg–Eggs fertilized within 10 min

Butts et al., Larvi 2013, poster

EGG AND LARVAL REARING TECHNIQUES

UNIVERSITEIT NTNU - Trondheim Norwegian University of Science and Technology



Establishment of biotechnical conditions for egg incubation and larval rearing

- Water quality management
- Recirculation, flow-through
- Mesocosmos etc.

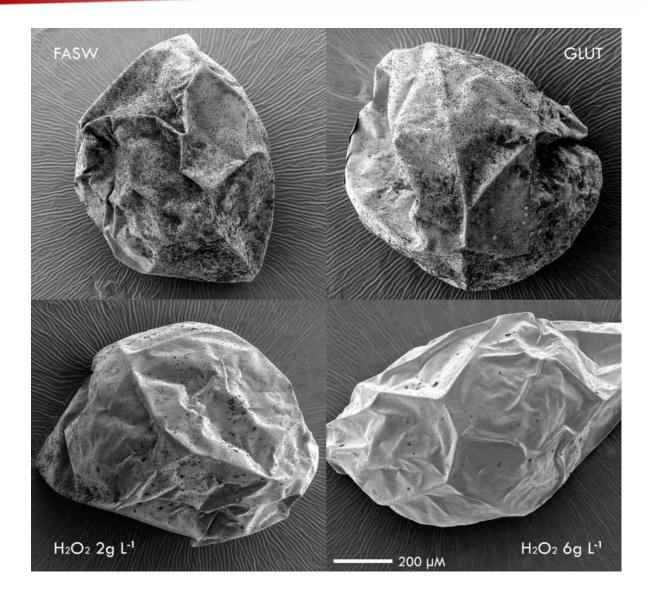


Transport and export

- Transport of eggs by flight from Denmark to Norway
- Successful parallel experiments

EGG AND LARVAL REARING TECHNIQUES

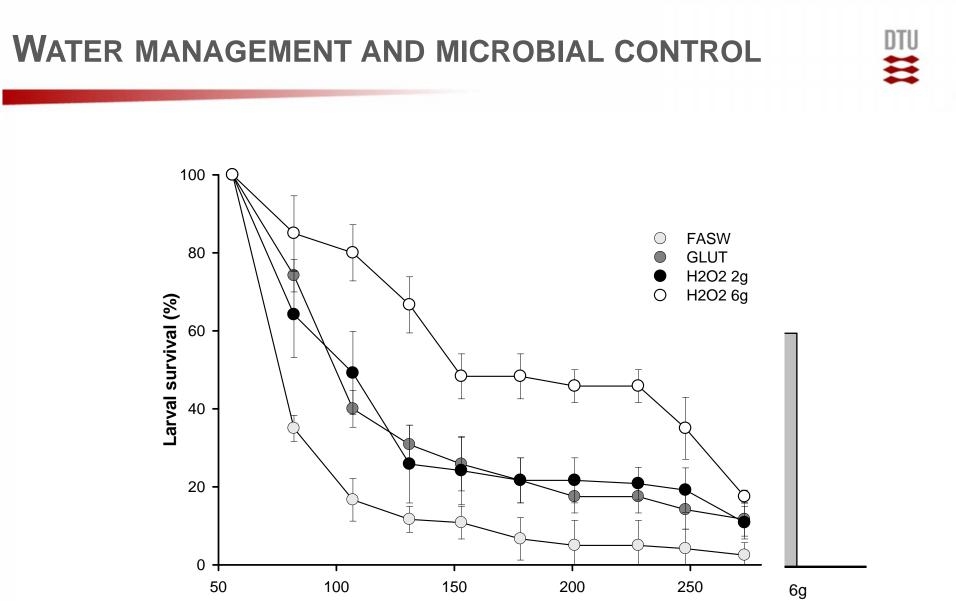




Incubation: Test of egg disinfection treatments

Treatments:

- Filtered autoclaved
 Seawater
- > Glutar aldehyde
- > Hydrogen peroxide
 - low 2 g L⁻¹
 - high 6 g L⁻¹



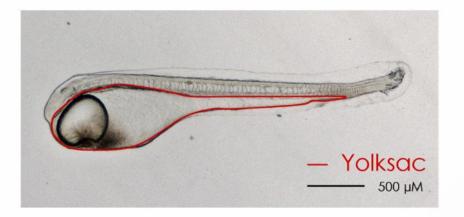
Hours after fertilization

Larval survival in relation to disinfection treatment

QUALITY OF YOLKSAC LARVAE

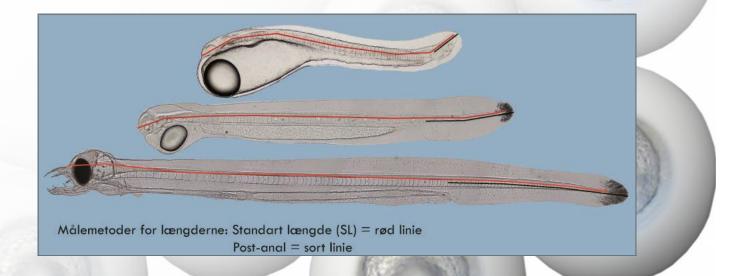


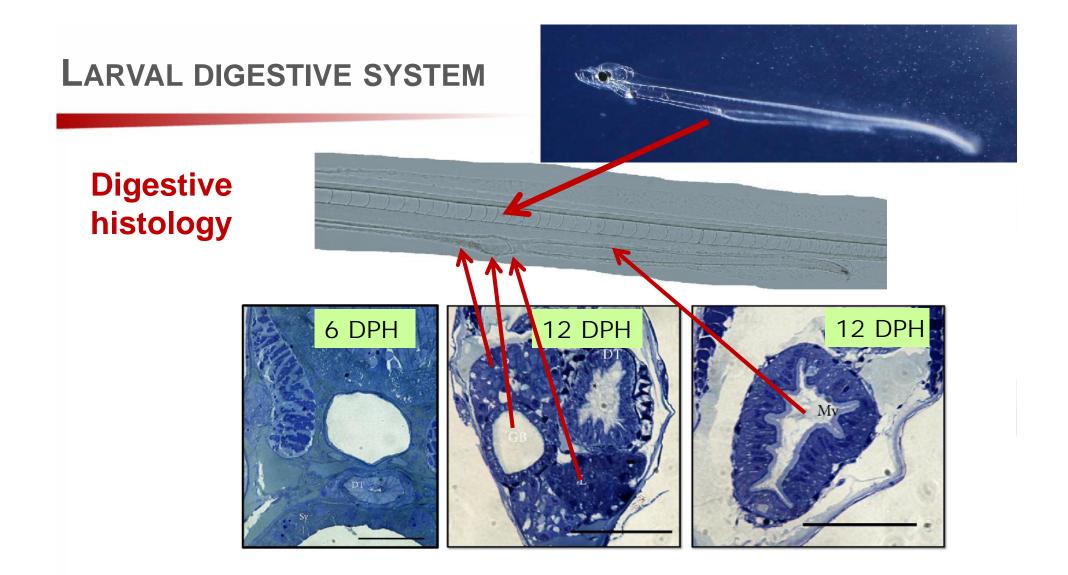
Measurments of morphological characteristics and general developmental success



Amount and use of yolk

Length increase

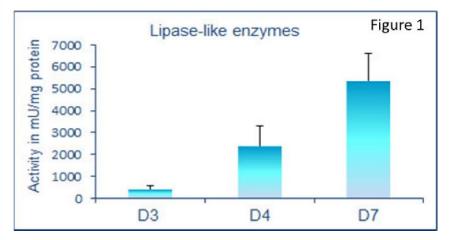




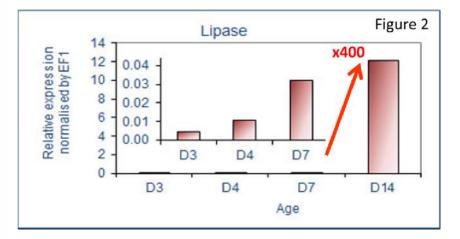
- Large yolk syncytium at 6 DPH,
- Relatively immature liver/pancreas at 12 DPH

LARVAL DIGESTIVE SYSTEM





Enzyme activity (biochemical study)



Enzyme gene expression

Digestive enzymes in eel larvae day 3-14

Very high activities in lipase-like enzymes after hatching

- Similar development in expression of the enzymes' corresponding coding genes
 - lipase, amylase, trypsin and aminopeptidase N

Data suggest that yolksac eel larvae have an elevated nutritional requirement for lipids

Important for development of new strategies for feeding eel larvae?

LARVAL FEEDING STAGE AND FEED?

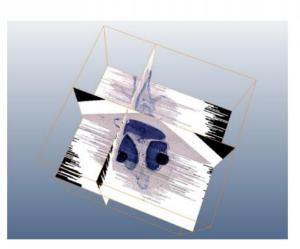
European eel larvae 14-15 DPH

What can they eat?

Analysis of larval head morphology

Study of larval behaviour







M. Bouillart et al., Lariv 2013

CONCLUSION



Successful results of PRO-EEL

- Enhanced broodstock feeds for high quality eggs and larvae comparable to wild broodstocks
- Optimal fertilization protocols
- Stable production of viable eggs and larvae
- ✓ Culture of yolksac larvae
- Insight in to larval nutritional requirements

Future goals

Identification of suitable feed, feeding larval culture and ongrowing